MICROBIAL CONVERSION OF MILBEMYCINS: 29-HYDROXYLATION OF MILBEMYCINS BY GENUS Syncephalastrum

KEIKO NAKAGAWA, KAZUO SATO[†], Yoshihisa Tsukamoto[†] and Akio Torikata

Fermentation Research Laboratories, Sankyo Co., Ltd., 1-2-58, Hiromachi, Shinagawa-ku, Tokyo 140, Japan [†]Agricultural Chemicals Research Laboratories, Sankyo Co., Ltd., 1041 Yasu, Yasu-cho, Shiga 520-23, Japan

(Received for publication October 4, 1991)

Milbemycins are a family of sixteen-membered macrolides produced by *Streptomyces hygroscopicus* subsp. *aureolacrimosus*, and demonstrate potent and broad spectrum activity as anthelmintics, acaricides and insecticides.^{$1 \sim 3$}

In previous papers, we reported that milbemycins were converted into 30-hydroxy, 13β -hydroxy, and 13β ,29-dihydroxy derivatives by using *Amycolata autotrophica* subsp. *amethystina* ATCC 35204,

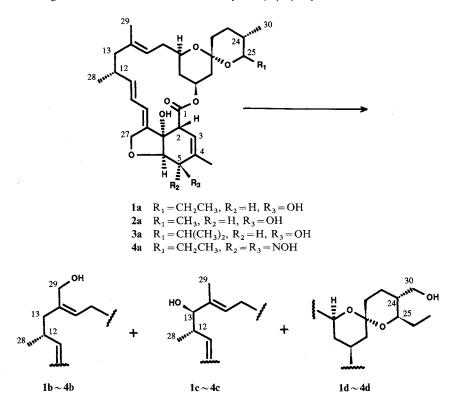
Cunninghamella echinulata ATCC 9244, and Streptomyces cavourensis SANK 67386, respectively.^{4~6)} As part of our continuing screening effort for microbial conversion of milbemycins, we observed that Syncephalastrum racemosum and Syncephalastrum nigricans had conversion abilities to milbemycin A_4 (1a).

The present paper describes the isolation and identification of converted products of milberrycin A_4 (1a), A_3 (2a), D (3a) and 5-ketomilberrycin A_4 5-oxime (4a) by *S. racemosum* (Fig. 1).

Milbemycins conversion were followed by TLC (Merck Art. No. 5715: EtOAc) and HPLC (column: Waters, Nova pak C_{18} 8 mm × 10 cm; solvent: system 1, acetonitrile - water (75:25), with a flow rate of 1.5 ml/minute; system 2, acetonitrile - water (55:45), with a flow rate of 1.0 ml/minute; detector: UV 243 nm).

S. racemosum SANK 42672 was cultured in five 500-ml Erlenmeyer flasks containing 100 ml MY medium composed of glucose 1.0%, Polypepton (Daigo Nutritive Chemicals) 0.5%, yeast extract (Difco) 0.3%, and malt extract (Difco) 0.3% (pH $6.3 \sim 6.5$) at 26°C on a rotary shaker (200 ~ 220 rpm).

Fig. 1. Microbial conversion of milbemycin A₄ by Syncephalastrum racemosum.



0.1.4.4	Concentration (µg/ml)	Conversion time (days)	Product ^a yield (%)		
Substrate			b	с	d
1a	500	7	5.7	1.5	2.8
2a	250	7	1.6	2.1	1.2
3a	500	7	1.0	1.1	1.0
4 a	500	7	2.1	3.5	1.9

Table 1. Conversion of milbemycins by Syncephalastrum racemosum SANK 42672.

^a **b**: 29-Hydroxy derivative; **c**: 13β -hydroxy derivative; **d**: 30-hydroxy derivative.

After the cultivation for 2 days, milberrycin A_4 (5% (w/v) in 1,4-dioxane) was added to a final concentration of 500 µg/ml and cultivation was additionally continued for 7 days. Then the culture broth was filtered and the filtrate was extracted with three 300-ml portions of EtOAc. EtOAc extract was dried over anhydrous sodium sulfate and evaporated. The mycelium was extracted with 80% MeOH. MeOH was then removed from the extract by evaporation and resulting aqueous solution was extracted and evaporated in a similar manner as the filtrate. These extracts were combined and evaporated in vacuo. The residue was subjected to silica gel chromatography ($20 \sim 90\%$ EtOAc in *n*-hexane as an eluent) and three products were isolated and characterized as 29-hydroxymilbemycin A₄ (1b), 13 β -hydroxymilbemycin A₄ (1c) and 30-hydroxymilberrycin A_4 (1d). These hydroxylated products were identical to the authentic compounds on the basis of the comparison of NMR and mass spectrum.^{4,5,7)} According to the patent procedures,⁷⁾ which was not easily accessible (multi steps and low yield), authentic 29-hydroxymilbemycin A4 was obtained with synthetic methods. In contrast to the synthetic procedures, preparation of 29-hydroxymilbemycin A₄ by a single microbial conversion step offers interesting and variable prospects.

Milbemycins A₃ (2a), D (3a) and 5-ketomilbemycin A₄ 5-oxime (4a)⁸⁾ were converted using similar method as for milbemycin A₄ (1a). The corresponding 29-hydroxy derivatives, 30- and 13 β -hydroxy derivatives were also obtained. Yields of each conversion are summarized in Table 1. The Rf values on TLC and HPLC retention times of 29-hydroxy derivatives are listed in Table 2. The physicochemical properties of 29-hydroxymilbemycins A₄ (1b), A₃ (2b), D (3b), and 29-hydroxy-5-ketomilbemycin A₄ (4b) were as follows:

29-Hydroxymilbemycin A₄ (**1b**): IR (KBr) cm⁻¹ 3650~3150 (br s), 2960 (s), 2928 (s), 2873 (s), 1715 (s); ¹H NMR (270 MHz, CDCl₃) δ 5.70~5.85 (2H, m, 9-H, 10-H), 5.31~5.48 (3H, m, 3-H, 11-H, 19-H),

Table 2. TLC Rf values and HPLC retention times of milbemycins and conversion products.

Compounds	TLC Rf ^ь	HPLC Rt's ^b (minutes)		
Compound ^a	values	System 1	System 2	
1a	0.59	16.07	·	
1b	0.38	3.54	11.50	
1c	0.46	3.50	10.86	
1d	0.44	3.08	8.91	
2a	0.59	11.80		
2b	0.38	3.02	8.27	
2c	0.46	3.02	8.04	
2d	0.41	2.83	7.12	
3a	0.62	24.64		
3b	0.39	4.74	19.58	
3c	0.48	4.59	18.26	
3d	0.50	3.64	12.65	
4 a	0.69	18.91	—	
4b	0.60	3.86	15.32	
4 c	0.60	3.84	14.75	
4d	0.55	3.22	10.65	

^a a: Substrate; **b**~**d**: products.

^b Rf values and retention times relative to 1c.

5.12 (1H, t, J=7.9 Hz, 15-H), 4.70, 4.64 (2H, ABq, J=15.3 Hz, 27-H₂), 4.29 (1H, br s, 5-H), 4.27, 3.94 (2H, ABq, J=12.1 Hz, 29-H₂), 4.11 (1H, s, 7-OH), 3.96 (1H, d, J=6.0 Hz, 6-H), 3.60 (1H, m, 17-H), 3.25 (1H, m, 2-H), 3.07 (1H, dt, $J_d=2.6$ Hz, $J_t=9.4$ Hz, 25-H), 2.49 ~ 2.63 (2H, m, 13-H, 16-H), 2.17 ~ 2.40 (3H, m, 5-OH, 12-H, 16-H), 2.00 (1H, ddd, J=1.7, 4.9, 12.1 Hz, 20-H), 1.87 (3H, s, 26-H₃), 1.20 ~ 1.85 (10H, m, 13-H, 18-H, 20-H, 22-H₂, 23-H₂, 24-H, 31-H₂), 1.03 (3H, d, J=6.9 Hz, 28-H₃), 0.99 (3H, t, J=6.9 Hz, 32-H₃), 0.80 ~ 0.95 (1H, m, 18-H), 0.83 (3H, d, J=6.5 Hz, 30-H₃); MS m/z 558 (M⁺, C₃₂H₄₆O₈), 540, 430, 412, 394, 369, 330, 279, 261, 246, 195, 181, 167, 151; HREI-MS calcd for C₃₂H₄₆O₈: 558.3193, found: 558.3193.

29-Hydroxymilbemycin A₃ (**2b**): IR (KBr) cm⁻¹ 3650 ~ 3150 (br s), 2967 (s), 2928 (s), 2874 (s), 1716 (s); ¹H NMR (270 MHz, CDCl₃) δ 5.70 ~ 5.82 (2H, m, 9-H, 10-H), 5.31 ~ 5.47 (3H, m, 3-H, 11-H, 19-H), 5.15 (1H, t, J = 7.8 Hz, 15-H), 4.70, 4.64 (2H, ABq, J = 15.3 Hz, 27-H₂), 4.29 (1H, m, 5-H), 4.27, 3.94 (2H, ABq, J = 12.3 Hz, 29-H₂), 4.10 (1H, s, 7-OH), 3.96 (1H, d, J = 6.4 Hz, 6-H), 3.58 (1H, m, 17-H), 3.20 ~ 3.31 (2H, m, 2-H, 25-H), 2.50 ~ 2.69 (2H, m, 13-H, 16-H), 2.22 ~ 2.40 (3H, m, 5-OH, 12-H, 16-H), 2.00 (1H, ddd, J = 1.6, 4.9, 12.1 Hz, 20-H), 1.87 (3H, d, J = 1.6 Hz, 26-H₃), 1.15 ~ 1.80 (8H, m, 13-H, 18-H, 20-H, 22-H₂, 23-H₂, 24-H), 1.15 (3H, d, J = 6.1 Hz, 31-H₃), 1.03 (3H, d, J = 6.5 Hz, 28-H₃), 0.80 ~ 0.95 (1H, m, 18-H), 0.83 (3H, d, J = 6.5 Hz, 30-H₃); MS m/z 544 (M⁺, C₃₁H₄₄O₈), 508, 458, 416, 398, 351, 319, 285, 266, 247, 215, 181, 163, 153; HREI-MS calcd for C₃₁H₄₄O₈: 544.3036, found: 544.3023.

29-Hydroxymilbemycin D (3b): IR (KBr) cm^{-1} 3650~3150 (br s), 2962 (s), 2928 (s), 2872 (s), 1714 (s); ¹H NMR (270 MHz, CDCl₃) δ 5.70 ~ 5.85 (2H, m, 9-H, 10-H), 5.27 ~ 5.46 (3H, m, 3-H, 11-H, 19-H), 5.11 (1H, dd, J = 6.3, 9.7 Hz, 15-H), 4.67 (2H, br s, $27-H_2$, 4.29 (1H, d, J = 6.4 Hz, 5-H), 4.26, 3.94 (2H, ABq, J = 12.0 Hz, 29-H₂), 4.05 (1H, br s, 7-OH), 3.96 (1H, d, J = 6.4 Hz, 6-H), 3.65 (1H, m, 17-H),3:26 (1H, m, 2-H), 3.07 (1H, dd, J=2.0, 9.3 Hz, 25-H), 2.48~2.64 (2H, m, 12-H, 16-H), 2.25~2.38 (2H, m, 16-H, 13-H), 2.00 (1H, ddd, J=1.5, 4.9)11.7 Hz, 20-H), 1.87 (3H, s, 26-H₃), 1.25~1.95 (9H, m, 13-H, 18-H, 20-H, 22-H₂, 23-H₂, 24-H, 31-H), 1.05 (3H, d, J = 6.8 Hz, 32-H₃), 1.03 (3H, d, $J = 6.4 \text{ Hz}, 28 \text{-H}_3$, 0.75 ~ 0.90 (1H, m, 18-H), 0.86 $(3H, d, J=6.8 Hz, 33-H_3), 0.80 (3H, d, J=6.9 Hz,$ $30-H_3$; MS m/z 572 (M⁺, C₃₃H₄₈O₈) 474, 444, 408, 330, 275, 209, 151; HREI-MS calcd for C33H48O8: 572.3349, found: 572.3329.

29-Hydroxy-5-ketomilbemycin A_4 5-oxime (4b): IR (KBr) cm⁻¹ $3650 \sim 3150$ (br s), 2959 (s), 2928 (s), 2872 (s), 1715 (s); ¹H NMR (270 MHz, CDCl₃) δ 5.73 ~ 5.91 (3H, m, 3-H, 9-H, 10-H), 5.31 ~ 5.51 (2H, m, 11-H, 19-H), 5.13 (1H, dd, J=7.2, 15.1 Hz, 15-H), 4.77, 4.69 (2H, dABq, $J_d = 2.2 \text{ Hz}$, $J_{AB_d} =$ 14.7 Hz, 27-H₂), 4.67 (1H, s, 6-H), 4.27, 3.94 (2H, AB_q , J = 11.7 Hz, 29-H₂), 4.00 (1H, s, 7-OH), 3.60 (1H, m, 17-H), 3.38 (1H, m, 2-H), 3.07 (1H, dt, $J_{\rm d} = 1.7 \,{\rm Hz}, J_{\rm t} = 8.5 \,{\rm Hz}, 25 \,{\rm H}), 2.49 \,{\sim}\, 2.63 \,(2 {\rm H}, {\rm m})$ 12-H, 16-H), 2.15~2.36 (2H, m, 13-H, 16-H), 2.00 (1H, ddd, J=2.0, 5.3, 12.1 Hz, 20-H), 1.94 (3H, d, d) $J = 2.0 \text{ Hz}, 26 \text{-H}_3$, 1.20 ~ 1.85 (10H, m, 13-H, 18-H, 20-H, 22-H₂, 23-H₂, 24-H, 31-H₂), 1.03 (3H, d, $J = 6.4 \text{ Hz}, 28 \text{-H}_3), 1.00 (3\text{H}, \text{t}, J = 7.3 \text{ Hz}, 32 \text{-H}_3),$ $0.80 \sim 0.95$ (1H, m, 18-H), 0.82 (3H, d, J = 6.3 Hz, 30-H₃); MS m/z 571 (M⁺, C₃₂H₄₅NO₈), 537, 429, 397, 370, 336, 292, 279, 240, 211, 195, 181, 167, 149; HREI-MS calcd for C₃₂H₄₅NO₈: 571.3145, found:

Table 3. 29-Hydroxymilbemycin A_4 -producing strains from milbemycin A_4 .

Microorganism	Conversion efficiency ^a		
	1b	1c	1d ^b
Syncephalastrum racemosum SANK 41872	+2	+1	+1
S. racemosum SANK 41972	+1	+1	tr
S. racemosum SANK 42672	+2	+1	tr
S. nigricans SANK 42172	+2	+1	+1
S. nigricans SANK 42272	+1	+1	tr
S. nigricans SANK 42372	+1	+1	+1
Rhizopus circinans SANK 16480	+1	+2	tr

^a +1: $0.5 \sim 10\%$, +2: $10 \sim 30\%$, tr: trace (HPLC analysis).

^b 1b: 29-Hydroxymilbemycin A₄, 1c: 13β-hydroxymilbemycin A₄, 1d: 30-hydroxymilbemycin A₄.

571.3134.

Other cultures which were found to convert milberrycin A_4 (1a) to 29-hydroxymilberrycin A_4 (1b) and their conversion efficiency are given in Table 3. When milberrycin A_4 (1a) was hydroxylated to 13β ,29-dihydroxymilbemycin A₄ by Streptomyces cavourensis SANK 67386 (unpublished data), trace amounts of 29-hydroxymilbemycin A₄ (1b) was found in the fermentation mixtures by HPLC detection. In the current study, Syncephalastrum spp. afforded 29-hydroxymilbemycin A_4 (1b) as a major product from milbertycin A_4 (1a). In the previous report, only Syncephalastrum spp. could convert ML-236B,⁹⁾ a competitive inhibitor of 3-hydroxy-3methylglutaryl-CoA reductase, predominantly to 6a-hydroxy-ML-236B while a number of organisms hydroxylated at 6β -position.^{10,11} It is of interest to note that the difference of stereoselectivity in the hydroxylation of ML-236B between Syncephalastrum spp. and other microorganisms was one of site selectivity in the hydroxylation of milbemycins. The characterization of the oxygenase in Syncephalastrum spp. is now under investigation. It was also found that Rhizopus circinans, which similar to Syncephalastrum belongs to the Mucorales, possessed 29-hydroxylation activity of milbemycin A₄ (1a) but with lower activity.

In conclusion, the microbial conversion of milbemycins by *Syncephalastrum* spp. to effect the preferential hydroxylation at the 29-position was apparently indicated.

References

- TAKIGUCHI, Y.; H. MISHIMA, M. OKUDA, M. TERAO, A. AOKI & R. FUKUDA: Milbemycins, a new family of macrolide antibiotics: Fermentation, isolation and physico-chemical properties. J. Antibiotics 33: 1120~1127, 1980
- OKAZAKI, T.; M. ONO, A. AOKI & R. FUKUDA: Milbemycins, a new family of macrolide antibiotics: Producing organism and its mutants. J. Antibiotics 36: 438~441, 1983
- MISHIMA, H.; J. IDE, S. MURAMATSU & M. ONO: Milbemycins, a new family of macrolide antibiotics. Structure determination of milbemycins D, E, F, G, H, J and K. J. Antibiotics 36: 980~990, 1983
- NAKAGAWA, K.; A. TORIKATA, K. SATO & Y. TSUKAMOTO: Microbial conversion of milbemycins: 30-Oxidation of milbemycin A₄ and related compounds by *Amycolata autotrophica* and *Amycolatopsis mediterranei*. J. Antibiotics 43: 1321~1328, 1990
- 5) NAKAGAWA, K.; S. MIYAKOSHI, A. TORIKATA, K. SATO & Y. TSUKAMOTO: Microbial conversion of milbemycins: Hydroxylation of milbemycin A₄ and related compounds by *Cunninghamella echinulata* ATCC 9244. J. Antibiotics 44: 232~240, 1991
- 6) NAKAGAWA, K.; K. SATO, T. OKAZAKI & A.

TORIKATA: Microbial conversion of milbemycins: 13β ,29-Dihydroxylation of milbemycins by soil isolate *Streptomyces cavourensis*. J. Antibiotics 44: $803 \sim 805$, 1991

- GEHRET, J. C. (Ciba-Geigy A.-G.): Preparation of 29-(carbonyloxy)milbemycins for controlling parasites in animals and plants. Eur. Pat. Appl. 237 482, Mar. 6, 1986 [CA 110: 13497u, 1989]
- IDE, J.; S. MURAMATSU, Y. NAKADA & N. KITANO (Sankyo): Didehydromilbemycin derivatives. Eur. Pat. Appl. 110 677, Nov. 25, 1982 [CA 101: 210854m, 1984]
- ENDO, A.; M. KURODA & Y. TSUJITA: ML-236A, ML-236B, and ML-236C, new inhibitors of cholesterogenesis produced by *Penicillium citrinum*. J. Antibiotics 29: 1346~1348, 1976
- SERIZAWA, N.; K. NAKAGAWA, Y. TSUJITA, A. TERAHARA & H. KUWANO: 3α-Hydroxy-ML-236B (3α-hydroxycompactin), microbial transformation product of ML-236B (compactin). J. Antibiotics 36: 608~610, 1983
- 11) SERIZAWA, N.; S. SERIZAWA, K. NAKAGAWA, K. FURUYA, T. OKAZAKI & A. TERAHARA: Microbial hydroxylation of ML-236B (compactin). Studies on microorganisms capable of 3β -hydroxylation of ML-236B. J. Antibiotics 36: 887~891, 1983